HO CHI MINH CITY UNIVERSITY OF EDUCATION
JOURNAL OF SCIENCE

Vol. 18, No. 3 (2021): 425-430

Website: http://journal.hcmue.edu.vn

Research Article

STUDY ON CHEMICAL CONSTITUENTS OF THE LEAVES OF STERCULIA FOETIDA LINN.

Nguyen Thi Quynh Trang¹, Duong Thuc Huy², Pham Nguyen Kim Tuyen^{1*}

¹Saigon University, Vietnam

²Ho Chi Minh City University of Education, Việt Nam

*Corresponding author: Pham Nguyen Kim Tuyen – Email: phngktuyen@gmail.com Received: January 18, 2021; Revised: March 23, 2021; Accepted: March 25, 2021

ABSTRACT

1859-3100

Phytochemical data of Sterculia foetida Linn. are scarce. The leaves of Sterculia foetida Linn. collected in Binh Thuan Province were chemically investigated using multiple chromatographic methods. Three compounds, hesperidin (1), kaempferol (2) and ursolic acid (3) were isolated and elucidated. Their chemical structures were elucidated by comparing their spectroscopic data with those in previous studies. These compounds were found for the first time from the leaves of Sterculia foetida Linn. Compounds 1 was obtained for the first time in Sterculia genus.

Keywords: hesperidin; kaempferol; Sterculia foetida Linn.; ursolic acid

1. Introduction

Sterculia foetida Linn. (Sterculiaceae) is grown in tropical areas around the world (Chi, 2002; Peng et al., 2009). The latex of this species can be used as herbal drinks and beverages (Vo, 2002). The *S. foetida* extracts showed antidiabetic, anticancer, antibacterial, anti-inflammatory and analgesic activities (Peng et al., 2009). Flavonoids, triterpenoids, steroids and fatty acids were addressed (Peng et al., 2009; Mujumdar et al., 2000; Anjaneyulu et al., 1981, Kale et al., 2011). Our previous phytochemical studies on this species reported some oleanane-type triterpenoids, quercetin derivatives and phenolic compounds (Pham et al., 2018; Pham et al., 2019). This paper presented the isolation and structural elucidation of three compounds, including hesperidin (1), kaempferol (2) and ursolic acid (3) from *S. foetida* leaves collected in Binh Thuan Province, Vietnam.

Cite this article as: Nguyen Thi Quynh Trang, Duong Thuc Huy, & Pham Nguyen Kim Tuyen (2021). Study on chemical constituents of the leaves of *sterculia foetida* Linn. Ho Chi Minh City University of Education Journal of Science, 18(3), 425-430.

Figure 1. Chemical structures of isolated compounds 1-3

2. Experimental

2.1. General experimental procedures

The NMR spectra were recorded on a Bruker Avance 500 spectrometer (500 MHz for 1 H–NMR and 125 MHz for 13 C–NMR) in acetone- d_6 , and DMSO- d_6 solutions. Thin layer chromatography was carried out on silica gel 60 (Merck, 40-63 μ m) and spots were visualized by spraying with 10% H₂SO₄ solution, followed by heating.

2.2. Plant material

The leaves of *S. foetida* were collected in Binh Thuan Province, Vietnam in October 2017. Its scientific name was identified by botanist Dr. Dang Van Son, Institute of Tropical Biology. A voucher specimen (No.SFC/TUYEN-1017A) was deposited at the herbarium in the laboratory of Faculty of Environmental Science, Saigon University.

2.3. Extraction and isolation

The powdered leaves of *S. foetida* (12.0 kg) were macerated with ethanol for three times (3 x 40 L) at room temperature. The solvent was removed from the residue under low pressure to obtain the crude extract (1830 g). This crude extract was successively applied to liquid-liquid partition giving hexane (450.5 g), chloroform (650.0 g), ethyl acetate (30.1 g) extracts and the aqueous partition.

The chloroform extract (400.0 g) was applied on a silica gel column chromatography with mobile phase hexane-ethyl acetate (100:0, 75:25, 50:50, 25:75, 0:100, v/v, respectively) and then ethyl acetate-methanol (90:10, 80:20, 70:30, 60:40, 0:100, v/v, respectively) to give eight fractions (SFC.I-SFC.VIII). Fraction SFC.VII (9.2 g) was subjected to a silica gel column chromatography with gradient solvent of chloroform-methanol (80:20, 70:30, 60:40) to yield nine sub-fractions (SFC.VII.1-SFC.VII.9). Sub-fraction SFC.VII.1 (2.3 g) was chromatographed on a silica gel using chloroform-methanol (20:1, 10:1, 5:1) as eluent to give **3** (4.9 mg). Sub-fractions SFC.VII.6 (1.6 g) was rechromatographed on a silica gel eluting with chloroform-methanol (95:5, 90:10, 85:15, 80:20, 75:25) to afford **1** (5.2 mg) and **2** (7.8 mg).

• **Hesperidin** (**1**) (Lahmer et al., 2015). Yellow amorphous powder. The 1 H-NMR data (500 MHz, DMSO- d_6 , δ ppm, J in Hertz): 12.05 (1H, brs,5-OH), 6.95 (1H, dd, 8.0, 2.0, H-6'), 6.89 (1H, d, 2.0, H-5'), 6.81 (1H, d, 8.0, H-2'), 6.15 (1H, d, 2.0, H-6), 6.13 (1H, d, 2.0, H-8), 5.49 (1H, dd, 11.0, 5.0, H-2), 4.98 (1H, d, 7.5, H-1"), 4.57 (1H, brs, H-1"), 3.79 (3H, s, 4'-OCH₃), 3.23 (1H, m, H-3_{ax}), 2.77 (1H, d, 17.0, H-3_{eq}) and 1.09 (3H, d, 5.5,

H-6").¹³C-NMR (125 MHz, DMSO- d_6): Hesperitin skeleton: 78.1 (C-2), 42.3 (C-3), 197.8 (C-4), 163.0 (C-5), 96.4 (C-6), 165.3 (C-7), 95.6 (C-8), 162.6 (C-9), 103.5 (C-10), 131.1 (C-1'), 114.0 (C-2'), 146.2 (C-3'), 148.2 (C-4'), 112.0 (C-5'), 117.7 (C-6'); D-glucopyranosyl unit: 99.8 (C-1"), 72.0 (C-2"), 75.5 (C-3"), 70.7 (C-4"), 76.3 (C-5") and 66.0 (C-6"); L-rhamnopyranosyl moiety: 100.1 (C-1"'), 69.7 (C-2"'), 70.4 (C-3"'), 73.1 (C-4"'), 68.3 (C-5"') and 18.1 (C-6"'); and 55.8 (4'-OCH₃).

- **Kaempferol** (**2**) (Li et al., 2008). Yellow amorphous powder. The 1 H–NMR data (Acetone- d_6 , δ ppm, J in Hertz): 8.14 (2H, d, 8.5, H-2', H-6'), 7.13 (2H, d, 8.5, H-3', H-5'), 6.53 (1H, d, 2.0, H-8) and 6.26 (1H, d, 2.0, H-6). 13 C-NMR (125 MHz, Acetone- d_6): 147.0 (C-2), 137.1 (C-3), 176.6 (C-4), 162.3 (C-5), 99.3 (C-6), 166.1 (C-7), 94.5 (C-8), 158.2 (C-9), 104.5 (C-10), 123.6 (C-1'), 130.7 (C-2', C-6'), 116.1 (C-3', C-5') and 160.1 (C-4').
- Ursolic acid (3) (Silva et al., 2008). White powder. The ¹H-NMR data (DMSO-*d*₆, δ ppm, *J* in Hertz): 5.14 (1H, *t*, 4.0.;3.5, H-12), 3.20 (1H, *dd*, 8.0;7.0, H-3), 1.09 (3H, *s*, H-27), 0.98 (3H, *s*, H-25), 0.94 (3H, *s*, H-30), 0.92 (3H, *s*, H-26), 0.86 (3H, *s*, H-29), 0.81(3H, *s*, H-24) and 0.77 (3H, *s*, H-23). The ¹³C-NMR (125 HZ, DMSO-*d*₆): 38.6 (C-1), 26.9 (C-2), 78.8 (C-3), 36.9 (C-4), 55.2 (C-5), 18.3 (C-6), 31.0 (C-7), 39.4 (C-8), 47.5 (C-9), 36.9 (C-10), 23.2 (C-11), 125.5 (C-12), 138.1 (C-13), 41.9 (C-14), 32.5 (C-15), 22.1 (C-16), 47.7 (C-17), 52.7 (C-18), 30.0 (C-19), 38.8 (C-20), 27.8 (C-21), 36.7 (C-22), 27.9 (C-23), 15.9 (C-24), 15.5 (C-25), 16.8 (C-26), 23.4 (C27), 180.2 (C-28), 16.9 (C-29) and 21.0 (C-30).

3. Results and discussion

Compound 1 was obtained as a yellow amorphous powder. The ¹H-NMR spectrum of 1 displayed signals of a flavanone skeleton. Its spectrum showed a down field signal at δ 12.05 (1H, brs), indicating the presence of a chelated hydroxyl group at C-5 position. Two meta-coupled doublet proton signals at $\delta_{\rm H}$ 6.15 (1H, d, 2.0, H-6) and 6.13 (1H, d, 2.0, H-8), each integrated for one proton, were assigned to H-6 and H-8, respectively, of a 5,7dihydroxy A ring system. The 1',3',4'-trisubstituted B ring system in flavanone skeleton, in addition, was determined by the presence of three aromatic proton signals on ABX system at $\delta_{\rm H}$ 6.81 (1H, d, 8.0, H-2'), 6.89 (1H, d, 2.0, H-5') and 6.95 (1H, dd, 8.0, 2.0, H-6'). Furthermore, the flavanone skeleton was identified by the presence of one oxymethine proton at $\delta_{\rm H}$ 5.49 (1H, dd, 11.0, 5.0, H-2) and two methylen protons at $\delta_{\rm H}$ 3.23 (1H, m, H- 3_{ax}) and 2.77 (1H, d, 16.0, H- 3_{eq}). These data indicated that 1 was a hesperitin derivative. Moreover, the ¹H-NMR data displayed two anomeric proton signals at $\delta_{\rm H}$ 4.98 (1H, d, 7.5, H-1") and 4.57 (1H, brs, H-1"), which indicated 1 was a hesperitin diglycoside flavanone. The ¹³C-NMR spectra were fully supported by the presence of twenty-eight carbon signals, including fifteen carbons of a flavanone skeleton, twelve carbons of two sugar units and one methoxy carbon. The hesperitin flavanone derivative was confirmed by the presence of fifteen carbons, including one carbonyl carbon at δ_C 197.8 (C-4), twelve aromatic carbon signals from 95.6 ppm to 165.2 ppm, one oxymethine carbon at δ_C 78.1(C-2), and one methylen carbon at δ_C 42.3 (C-3). The β -D-glucopyranosyl unit was demonstrated by the displaying one anomeric proton with the large coupling constant at $\delta_{\rm H}$ 4.98 (1H, d, 7.5, H– 1") in the ${}^{1}H$ -NMR spectra and the addition of one anomeric carbon signal at $\delta_{\rm C}$ 99.8 (C-1") and five oxygenated carbons from 66.0 ppm to 76.3 ppm in the ¹³C-NMR spectrum. The α -L-rhamnopyranosyl moiety was identified by the presence of one singlet anomeric proton at δ_H 4.57 (1H, s, H–1") along with one anomeric carbon signal at δ_C 100.1 (C– 1"'), four oxygenated carbons from 18.1 ppm to 73.1 ppm and one typical Lrhamnopyranosyl methyl carbon at δc 18.1 (C-6"). HMBC correlations from aromatic protons at δ_H 6.81 (1H, d, 8.0, H-2'), and 6.95 (1H, dd, 8.0, 2.0, H-6') to carbon at δ_C 78.1 (C-2) confirmed the attachment of B benzene ring to C-2. Additionally, the methoxy proton signal at δ_H 3.79 (3H, s, 4'-OCH₃) correlated with aromatic carbon at δ_C 148.2 (C-4') in the HMBC spectra which confirmed that the aglycone was herperetin skeleton. The β -D-glucopyranosyl unit was attached to C-7 of a herperetin skeleton by the HMBC correlation between the anomeric proton at $\delta_{\rm H}$ 4.98 (1H, d, 7.5, H–1") and oxygenated carbon at $\delta_{\rm C}$ 165.3 (C-7). The α -L-rhamnopyranosyl moiety was linked to C-6 of the β -Dglucopyranosyl unit by the HMBC correlations between the anomeric proton $\delta_{\rm H}$ 4.57 (1H, s, H-1") and oxygenated methylene carbon at δ_C 66.0 (C-6"). The other COSY, HSQC and HMBC correlations were strongly agreed with the assignment. Based on these spectroscopic data and comparison with previous report (Lahmer et al., 2015), the chemical structure of **1** was determined as hesperitin 7-O-[α -L-rhamnopyranosyl-(1 \rightarrow 6)]- β -D-glucopyranoside (Hesperidin). Hesperidin was isolated for the first time from Sterculia genus.

$$H_3C$$
 H_3C
 H_3C
 H_4C
 H_5
 H_5
 H_5
 H_5
 H_7
 H_7

Figure 2. The main HMBC correlations of compound 1

Compound **2** was obtained as a yellow amorphous powder. Its NMR spectrum showed similar signals of **1**, except for the appearing of one oxygenated aromatic carbon at $\delta_{\rm C}$ 147.0 (C-2), one olefinic carbon at $\delta_{\rm C}$ 137.1 (C-3), and disappearing two sugar units in **2** which was corresponded to a flavonol framework. The *para*–disubstituted benzene ring (ring B) was determined by the displaying of two doublet proton signals with a large coupling constant at $\delta_{\rm H}$ 8.14 (2H, d, 8.5 Hz, H–2', H–6') and 7.13 (2H, d, 8.5 Hz, H–3', H–5'). These data indicated that **2** was a kaempferol skeleton. The ¹³C–NMR spectrum

were strongly supported by the displaying of 15 carbons in the down field, comprising one carbonyl carbon at δ_C 176.6 (C–4), and the rest fourteen carbons from 94.5 ppm to 162.3 ppm. The good compatibility between its NMR data and those in the literature (Li et al.., 2008) determined the structure of 4 to be kaempferol.

Compound **3** was obtained as a white powder. The NMR data of **3** displayed signals of a triterpenoid skeleton. The 1 H-NMR determined the signals of an olefinic proton at $\delta_{\rm H}$ 5.14 (1H, t, 4.0.;3.5, H-12), an oxygenated proton at $\delta_{\rm H}$ 3.20 (1H, dd, 8.0;7.0, H-3) and seven methyl proton signals from 0.77 ppm to 1.09 ppm. The 13 C-NMR exhibited thirty carbons, including one carboxyl carbon at $\delta_{\rm C}$ 180.2 (C-28), two olefinic carbons at $\delta_{\rm C}$ 125.5 (C-12), and 138.1 (C-13) of a double bond at C-12/C-13 and the rest carbons in ursane skeleton. The HSQC and HMBC spectra determined this suggestion. The comparison NMR data of **3** with those reported in the literature, **3** were assigned as ursolic acid (Silva et al., 2008)

4. Conclusions

From the leaves of *S. foetida* collected in Binh Thuan Province, three compounds, including one diglycoside flavanone hesperidin (1), one flavonol kaempferol (2) and one ursane-type triterpenoid ursolic acid (3) were isolated for the first time. Their chemical structures were determined by using the NMR spectroscopic method as well as comparison with previous studies. Compound 1, to the best of our knowledge, was isolated from *Sterculia* genus for the first time. Further studies on this species are on the progress.

Conflict of Interest: Authors have no conflict of interest to declare.

REFERENCES

- Anjaneyulu, A. S. R., & Murty, V. S. (1981). Two rare tetramethyl ethers of quercetin from *Sterculia foetida* Linn. *Indian Journal of Chemiscal Section B*, 20(1), 87-88.
- Kale, S. S., Darade, V., & Thakur, H. A. (2011). Analysis of fixed oil from *Sterculia foetida* Linn. *International Journal of Pharmaceutical Sciences and Research*, 2(11), 2908-2014.
- Lahmer, N., Belboukhari, N., Cheriti, A., & Sekkoum, K. (2015). Hesperidin and hesperitin preparation and purification from *Citrus sinensis* peels. *Der Pharma Chemica*, 7(2), 1-4.
- Li, Y. L., Li, J., Wang, N. L., & Yao, X. S. (2008). Flavonoids and a new polyacetylene from *Bidens parviflora* Willd. *Molecules*, 13(8), 1931-1941.
- Mujumdar, A. M., Naik, D. G., Waghole, R.J., Kulkarni, D. K., & Kumbhojkar, M. S. (2000). Pharmacological studies on *Sterculia foetida* leaves. *Pharmaceutical Biology*, *38*(1), 13-17.
- Peng, F. X., Feng, Zi, M. F., Ya, N. Y., Pei, C. Z. (2009). Two flavonoid glycosides and a phenylpropanoid glucose ester from the leaves of *Sterculia foetida*. *Journal of Asian Natural Products Research*, 11(8), 766-771.

- Pham, D. T., Doan, T. D. C., Nguyen, T. P., Mai, D. T., Pham, N. K. T., & Nguyen, K. P. P. (2018). Quercetin derivatives from the ethyl acetate of the leaves of *Sterculia foetida* Linn. *Vietnam Journal of Chemistry*, 56(4e), 120–123.
- Pham, N. K. T, Nguyen, T. D., Tran, D. C. D., Tuan, H. D., Nguyen, T. M. A., Trong, D. T., Mai, D. T., & Nguyen, T. P. (2019). Stercufoetin A, new oleanane-type triterpenoid from the leaves of *Sterculia foetida* L. *Natural Product Research*, 1-6. doi.org/10.1080/14786419.2019.1644508, 2020.
- Silva, M. G. V., Vieira, Í. G. P., Mendes, F. N. P., Albuquerque, I. L., Dos Santos, R. N., Silva, F.O., & Morais, S. M. (2008). Variation of ursolic acid content in eight *Ocimum* species from northeastern Brazil. *Molecules*, *13*(10), 2482-2487.
- Vo, V. C. (2002). Dictionary of medical plants in Vietnam. Med Publishing House, 1083-1084.

NGHIÊN CỦU THÀNH PHẦN HÓA HỌC CỦA LÁ CÂY TRÔM STERCULIA FOETIDA LINN.

Nguyễn Thị Quỳnh Trang¹, Dương Thúc Huy², Phạm Nguyễn Kim Tuyến^{1*}

¹Trường Đại học Sài Gòn, Việt Nam

²Trường Đại học Sư phạm Thành phố Hồ Chí Minh, Việt Nam

*Tác giả liên hệ: Phạm Nguyễn Kim Tuyến – Email: phngktuyen@gmail.com
Ngày nhận bài: 18-01-2021; ngày nhận bài sửa: 23-3-2021, ngày chấp nhận đăng: 25-3-2021

TÓM TẮT

Nghiên cứu hóa thực vật của loài Sterculia foetida Linn. chưa được tìm thấy nhiều. Nghiên cứu được thực hiện trên lá cây Trôm Sterculia foetida Linn. thu hoạch ở tỉnh Bình thuận bằng các phương pháp sắc kí khác nhau. Ba hợp chất hesperidin (1), kaempferol (2) và ursolic acid (3) được cô lập và xác định cấu trúc hóa học. Cấu trúc hóa học của các hợp chất được xác định bằng các phương pháp phổ nghiệm đồng thời so sánh với các dữ liệu phổ đã được công bố. Đây là lần đầu tiên các hợp chất này được cô lập từ lá cây Trôm Sterculia foetida Linn. Hợp chất 1 lần đầu tiên được biết có hiện diện trong chi Sterculia.

Từ khóa: hesperidin; kaempferol; Sterculia foetida Linn.; ursolic acid