DETERMINATION OF AMMONIUM AND POTASSIUM EXTRACTED FROM SOIL SAMPLES BY CAPILLARY ELECTROPHORESIS WITH CAPACITIVELY COUPLED CONTACTLESS CONDUCTIVITY DETECTION (CE-C⁴D)

Đến tòa soạn 23 - 3 - 2016

Van Anh Nguyen

Department of Environmental Sciences and Technologies, Hanoi Metropolitan University

Thi Trang Do, Van Lau Ha, Hung Viet Duong, Tien Duc Pham, Thi Anh Huong Nguyen*

Faculty of Chemistry - Hanoi University of Science - Vietnam National University, Hanoi,

Thu Hien Le

Faculty of Transport Economics, University of Transport Technology

TÓM TẮT

XÁC ĐỊNH AMONI VÀ KALI TRONG DỊCH CHIẾT CỦA MẪU ĐẤT BẰNG PHƯƠNG PHÁP ĐIỆN DI MAO QUẨN SỬ DỤNG DETECTOR ĐỘ DẪN KHÔNG TIẾP XÚC (CE-C⁴D)

Bài báo nghiên cứu quy trình phân tích hai ion NH_4^+ và K^+ trong dịch chiết mẫu đất nông nghiệp bằng kĩ thuật điện di mao quản sử dụng detector độ dẫn không tiếp xúc. Các điều kiện phân tích đã được tối ưu bao gồm: Dung dịch đệm điện di sử dụng hỗn hợp đệm histidine/axit acetic (His/Ace) (pH = 4), 2mM 18-crown-6; mao quản Silica: đường kính ngoài (O.D.) 365 μ m, đường kính trong (I.D.) 50 μ m, chiều dài tổng L_t = 50cm và chiều dài hiệu dụng L_{eff} = 40cm; điện thế tách: 20kV; bơm mẫu theo phương pháp thủy động lực theo kiểu xiphong ở độ cao 15cm, trong thời gian 20s. Quy trình được áp dụng để phân tích dịch chiết 01 mẫu đất và 01 mẫu nước lấy tại cùng một khu canh tác nông nghiệp. Các kết quả phân tích được kiểm chứng bằng các phương

pháp quang phổ hấp thụ nguyên tử (kiểm chứng kết quả phân tích K⁺) và phương pháp quang phổ hấp thu phân tử (kiểm chứng kết quả phân tích NH₄⁺). Sai số tương đối giữa phương pháp phân tích và các phương pháp đo kiểm chứng đều dưới 15%. Kết quả cho thấy hàm lượng NH₄-N và K⁺ trong mẫu đất lần lượt là 29,6 và 1055 mg/kg đất khô. Hàm lượng NH₄-N và K⁺ trong mẫu nước lần lượt là 2,68 và 33,7 mg/L. Hàm lượng NH₄-N trong mẫu nước ruộng vượt hơn 5 lần quy chuẩn kĩ thuật Quốc gia về chất lượng nước mặt bảo vệ đời sống thủy sinh.

Key words: ammonium, potassium, contactless conductivity detector, agricultural soil

1. INTRODUCTION

Nitrogen and potassium are essential nutrients for plant growth, survival, development and reproduction. However, there are not always enough of these nutrients in the soil for a plant to grow healthily so that many farmers and gardeners use fertilizers as the nutrients for the soil. Despite of the fact that fertilizers are beneficial agriculture, excessive or improper fertilizer applying would induce to environmental and human health risks. The excess of nutrients in environment would cause soil pollution, surface eutrophication, groundwater water contamination... etc. Elevated levels of nutrients accumulated in crop products could make unexpected symptoms and diseases to consumers [1]. For better agricultural land use, it is necessary to quantify the nutrient residues that are available to be released to environment. The extractions of the exchangeable forms of K⁺ and NH₄⁺ can be performed by using salt solutions. Because NH₄⁺

 K^{+} and exhibit nearly identical common characteristics, extracting reagents for exchangeable potassium are ammonium salts. On the other hand, ammonium adsorbed on the exchange complex is normally removed by using potassium salts such as KCl or K₂SO₄ [2]. Thus, measurements of soil extracts for K⁺ and NH₄⁺ would be strongly interfered by the matrix effects. Capillary Electrophoresis (CE) with Capacitively Coupled Contactless Conductivity Detection (C⁴D) is a simple and inexpensive method that can applicable to simultaneously determine NH₄⁺ and K⁺ in aqueous samples. However, the separations of two cations solely based on differences in their electrophoretic mobility seem to be a challenge.

Numerous attempts were investigated to separate NH₄⁺ and K⁺ in aqueous solutions. One of the most successful approach was the employment of complex-forming reactions. Among comlexing agents, 18-crown-6 ether

was widely used to separate the NH₄⁺ and alkaline metal cations as it forms a stable complex with alkaline metal cations [3, 4, 5, 6, 7]. Adam J. Gaudry et al. separated an aqueous mixture comprising three inorganic cations (NH₄⁺, Na⁺ and Li⁺) in 50 mM acetic acid/10 mM L-histidine /2.5 mM18crown-6 ether electrolyte at pH 4.2. The limit of detections (LODs) of NH₄⁺, Na⁺, and Li⁺ were 1.54, 2.26, and 3.06mg/L, respectively [3]. Another separation of alkaline metal cations (K⁺, Na⁺, Li⁺) in aqueous solutions was obtained by using the background electrolyte (BGE) of 12 mM histidine adjusted to pH 4 with acetic acid and 2 mM 18-crown-6. The LOD for K⁺ was of 1.5 µM [8]. Solutions containing NH₄⁺, K⁺ and other cations were determined by Thanh Duc Mai et al. [5]. Background electrolyte solutions were His 12 mM (adjusted to pH 4 with acetic acid) in the presence of 2 mM of 18-crown-6; capillary was of 50 µm I.D., 36 cm effective length, and 50 cm total length, while separation voltage was 15 kV. The LODs were in the lower micromolar range and varied species. depending on Baseline separation between NH₄⁺ and K⁺ was achieved at the concentration of less than 100 µM for each cation [5]. Nevertheless, to our best knowledge, a simultaneous separation of NH₄⁺ and K⁺

aqueous solutions extracted from soils by CE-C⁴D has not been reported.

study, present operation the parameters were set up in our laboratory according to previously published papers for measuring NH₄⁺ and K⁺ in solutions. The optimizing aqueous of 18-crown-6 amount systematically investigated. Appropriate extracting solutions used to remove exchangeable faction of each ion from also examined. The soil were preliminary conditions were applied to detect NH₄⁺ and K⁺ in a water sample and the extracts of a soil sample collected from agricultural land in Vietnam.

2 MATERIALS AND METHODS

2.1. Materials

All the chemicals such as NH₄Cl, K_2SO_4 , His, $C_6H_9N_3O_2$, HCl, NaOH... etc were of analytical grade and purchased from Fluka or Merck. Ultrapure water system (Labconco, USA) with resistivity 18.2 M Ω was used to produce ultrapure water in all solutions and measurements. Stock solutions of ammonium and potassium daily prepared. were background electrolyte (BGE) solutions were prepared with the buffer of histidine and acetic acid (His/Ace) (pH=4) in the presence of 18-Crown-6. The pH of solutions was controlled by using an HI 2215 Hanna Instruments pH meter (Woonsocket, RI, USA). Fused silica capillaries of 50 μ m I.D. and 365 μ m O.D. with total length (L_t) of 50 cm and effective length (L_{eff}) of 40 cm (purchased from Polymicro, Phoenix, AZ, USA) were used for separations. The capillaries were preconditioned with 1 M NaOH for 10minutes and then, deionized water for

10minutes prior to flushing with buffer solutions. Experiments were performed using the portable semi-automated CE system supported by 3Sanalysis JSC. (http://www.3sanalysis.vn/) (Figure 1). More detailed information about the instrument was given elsewhere in T. A. H. Nguyen et. al. [9].



Figure 1. Portable semi-automated CE-C⁴D instrument

2.2. Methods

According to former references reviewed earlier, the BGE with the buffer adjusted to pH 4 was commonly used for the separations of the two cations using CE -C⁴D. In the present study, the buffer solution of His/Ace (pH=4) was selected. The high voltage

of 20kV was applied for separation. Hydrodynamic injection of samples was carried out by setting the high-voltage end of the capillary at 15cm height. The injection time was 20 seconds. Table 1 summarized the operating parameters used in this research.

*Table 1. Separation conditions of CE-C*⁴*D system for determine NH*₄⁺ and K^+ .

BGE composition	His/Ace buffer adjusted to pH 4	
	18-Crown-6	
High voltage	20kV	
Detection	C^4D	
Capillary	365 μm O.D., 50 μm I.D.; L_t = 50cm; L_{eff} =	
	40cm; Preconditioned	
Hydrodynamic injection	20s at 15cm height	

The BGE solutions with differences amount of 18-crown-6 were prepared in order to get optimum BGE composition for the separation of two cations. The concentrations of 18-crown-6 varied from 1 to 3mM. The concentrations of both NH₄⁺ and K⁺ were 0,1mM.

Appropriate soil extraction procedure investigated. Simulated was samples contaminated by 20mM of K⁺ and 2.2mM of NH₄⁺ were prepared by adding certain amount of K⁺ and NH₄⁺ solutions to uncontaminated soils. The samples were left for 1 day to reach to equilibrium, and then dried desiccator before further any preparation. For K⁺ extraction, the CH₃COONH₄ solutions of 0.01M and 0.1M were used. The ion NH₄⁺ was extracted with K₂SO₄ solutions of 0.001 and 0.1 M. The amount of 1.000 g dried soil and precisely 20ml of extracting agent were put into 50ml- falcon tubes and shaken for 1 hour using a shaker (Cole Parmer, 51704 Series). Finally, the mixture was centrifuged and filtered

through 0.45µm membrane. The filtrates were used for the K⁺ and NH₄⁺ measurements using CE-C⁴D. The levels of K⁺ and NH₄⁺ were compared with Flame - Atomic Absorption Spectroscopy (F-AAS) and Ultraviolet– Visible Spectroscopy ((UV-Vis), respectively.

One soil and one water samples were collected in agricultural regions Thuong Tin, Hanoi, Vietnam. The soil sample was immediately stored at 4°C and then dried in desiccator. The water sample was kept at 4°C and filtered through 0.45µm membrane before analysis.

3. RESULTS AND DISCUSSION

It was known that potassium forms a stable complex with 18-crown-6 [10]. Consequently, the presence of 18-crown-6 in the BGE increases the migration time of K⁺ while the migration time of NH₄⁺ stays unchanged. As clearly shown in Figure 2, the best separation was performed at the concentration of 2mM 18-crown-6. Lower concentrations than that of 2 mM

18-crown-6 would not be enough for reasonable separation of the two cations while higher concentrations seem to be not necessary. Thus, for further

experiments the BGE solutions contain Tris/Ace buffer (pH=4); 2 mM 18-Crown-6.

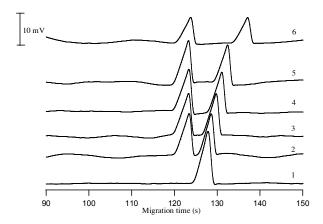


Figure 2. Separation of NH_4^+ and K^+ with BGE solutions containing different levels of 18-crown-6; Concentration of NH_4^+ and K^+ : 10^4M ; Concentration of 18-crown-6: (1): Non 18-crown-6 contained; (2) 1.0 mM; (3) 1.5 mM; (4) 2.0 mM; (5): 2.5 mM. (6): 3 mM

The calibration curves (six points) were extrapolated using standard addition method in order to avoid the matrix interference of the sample (Table 2). Linear range extended to the concentration of 200 μ M for each cations with a correlation coefficient (r²) of at least 0.99. The LOD and LOQ

values were calculated from peak areas corresponding to 3 and 10 times the baseline noise (S/N= 3 and S/N= 10), respectively. The LOD values (1.0 μ M for NH₄⁺ and 3.0 μ M for K⁺) are comparable to previously reported results [5, 8].

Table 2. Calibration curve extrapolated by standard addition method

Analyte	Range ^a (µM)	\mathbf{r}^2	LOD ^b (µM)	LOQ ^c (µM)
$\mathrm{NH_4}^+$	50.0-200	0.9988	1.0	5.0
K^{+}	50.0-200	0.9909	3.0	10.0

^a Six points

As can be seen in Table 3, the best extraction efficiency was achieved

when using 0.1M CH₃COONH₄ and 0.001M K₂SO₄ for extraction K⁺ and

^b S/N: 3

^c S/N: 10

NH₄⁺, respectively. Higher concentrations of extractants are unfavorable due to the strong matrix effect to the separation of the two

cations by CE, while lower ones are not enough to completely extract the exchangeable solutes.

Table 3. Extraction of the simulated soil sample for K^+ *and* NH_4^+

	Extractant	Extraction efficiency (%)
Extraction for K ⁺	CH ₃ COONH ₄ 0.01M	25.0
	CH ₃ COONH ₄ 0.1M	99.2
Extraction for NH ₄ ⁺	K ₂ SO ₄ 0.001M	109.0
	K_2SO_4 0.1M	_a

^aunable to detect

Figure 3 and 4 indicate good separation performances in the soil extract and water sample. Obtained results from CE-C⁴D are in good agreement with those from the confirmation method (F-AAS for K⁺ analysis and UV-Vis for NH₄⁺ analysis) (Table 4). This suggests that the conditions found are applicable for detection of NH₄⁺ and K⁺ in environmental samples. Levels of NH₄-N and K⁺ determined in the real soil sample were of 29.6 and 1055 mg/kg dried soil, respectively. Those highly

exceeded the normal level of NH₄-N (10 mg/kg) and the excessive level of K (> 800mg/kg) defined by D.A. Horneck et al. [11]. The concentration of NH₄-N in water sample was more than 5 times higher than the National technical regulation on surface water quality for protection of aquatic lives [12]. It should be noted that sufficiently high levels of NH₄⁺ and K⁺ in agricultural run-off and soil would cause nutrient imbalance and serious environmental problem.

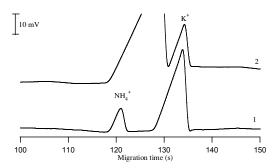


Figure 3. Separation of NH_4^+ and K^+ in the extracts of soil sample: (1) Soil extract for NH_4^+ ; (2) Soil extract for K^+ , dilution factor (DF) was 50 times

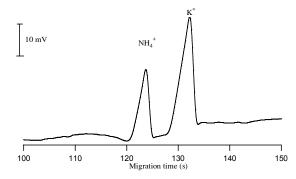


Figure 4. Simultaneous determination of NH_4^+ and K^+ in water sample (DF was 6 times)

Table 4. Concentrations of NH_4^+ *and* K^+ *in soil and water samples*

			-
Type of sample	Analyte	Concentration*	Confirmation
			(% difference)
Soil sample	\mathbf{K}^{+}	1055	5,7 ^a
	NH_4-N	29.6	10.8 ^b
Water sample	\mathbf{K}^{+}	33,7	12,9 ^a
	NH ₄ -N	2.68	0.50^{b}

^{*} mg/kg for soil sample; mg/L for water sample

4. CONCLUSIONS

conditions for simultaneous determination of NH₄⁺ and K⁺ by CE-C⁴D were optimized as follows: BGE: His/Ace buffer, 2 mM 18-crown-6; silica capillary: 365 µm O.D., 50 µm I.D.; $L_t = 50$ cm; $L_{eff} = 40$ cm; voltage: 20V; hydrodynamic injection: 20s at 15cm height. The procedure are applicable to examine NH₄⁺ and K⁺ in both soil and water samples. Excessive levels of NH₄⁺ and K⁺ were detected in soil. The concentration of NH₄⁺ in water sample was significantly higher than the National technical regulation on surface water quality for protection of aquatic

lives.

ACKNOWLEDGEMENT

This research was partly funded by Hanoi Metropolitan University under the project number C.2015-19.

REFERENCE

- [1] Stanley E. Manahan, (2009), Environmental chemistry, CRC Press.
- [2] Marc Pansu, Jacques Gautheyrou, (2006), Handbook of Soil Analysis: Mineralogical, Organic and Inorganic Methods, Springer.
- [3] Adam J. Gaudry, Michael C. Breadmore, and Rosanne M. Guijt (2013), In-plane alloy electrodes for

^a confirmation by F-AAS method

^b confirmation by UV-Vis method

- capacitively coupled contactless conductivity detection in poly (methylmethacrylate) electrophoretic chips, Electrophoresis, 34, 2980–2987. [4] Marko Stojkovic, Boris Schlensky, Peter C. Hauser, (2013), Referenced Capacitively Coupled Conductivity
- Detector for Capillary Electrophoresis, Electroanalysis, 25, No. 12, 2645 2650
- [5] Thanh Duc Mai, Thi Thanh Thuy Pham, Hung Viet Pham, Jorge Saiz, Carmen García Ruiz and Peter C. Hauser, (2013), Portable capillary electrophoresis instrument with automated injector and contactless conductivity detection, Anal. Chem., 85, 2333–2339
- [6] Thanh Duc Mai, Stefan Schmida, Beat Müller, Peter C. Hauser (2010), Capillary electrophoresis with contactless conductivity detection coupled to asequential injection analysis for manifold extended automated monitoring applications, Analytica Chimica Acta, 665, 1–6
- [7] Pavel Kubáň and Peter C. Hauser (2009), Ten years of axial capacitively coupled contactless conductivity detection for CZE a review. Electrophoresis, 30, pp.176-188.
- [8] Adam J. Gaudrya, Rosanne M. Guijtb, MirekMackaa, Joseph P. Hutchinsona, CameronJohnsa, Emily F. Hildera, Greg W. Dicinoski, Pavel

- N. Nesterenkoa, Paul R. Haddada, Michael C. Breadmorea, (2013), Online simultaneous and rapid separation of anions and cations from a single sample using dual-capillary sequential injection-capillary electrophoresis, Analytica Chimica Acta, 781, 80–87.
- [9] Thi Anh Huong Nguyen, Ngoc Mai Pham, Thi Tuoi Doan, Thi Thao Ta, Jorge Sáizc, Thi Quynh Hoa Nguyen, Peter C. Hauser, Thanh Duc Mai, (2014), Simple semi-automated portable capillary electrophoresis with contactless instrument conductivity detection for the determination of **β**-agonists pharmaceutical and pig-feed samples, Journal of Chromatography A, 1360, 305-311.
- [10] Harald Heiland, John A. Ringseth and Thorvald S. Brun (1979), Cation-Crown Ether Complex Formation in Water. II. Alkali and Alkaline Earth Cations and 12-Crown-4, 15-Crown-5, and 18-Crown-6, Journal of Solution Chemistry, Vol. 8, No. 11.
- [11] D. A. Horneck, D.M. Sullivan, J.S. Owen, and J.M. Hart (2011), Soil Test Interpretation Guide, EC 1478. Corvallis.
- [12] Ministry of Science, Technology and Environment (2011) QCVN 38:2011/BTNMT, National technical regulation on surface water quality for protection of aquatic lives.