

FABRICATION AND SURFACE MODIFICATION OF PT NANOWIRES FOR GLUCOSE DETECTION

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ABSTRACT: *In this paper we present a new fabrication technique that only uses conventional techniques of microtechnology such as microlithography, thin-film deposition and directional ion beam etching to make very narrow, wafer-scale length platinum (Pt) nanowires, named deposition and etching under angles (DEA). Then fabricated Pt nanowires electrodes were modified by using several chemicals to immobilize glucose oxidase (GOD) enzyme for application in glucose detection. A cyclic voltammetry (CV) technique was used to determine glucose concentrations. The detection results showed that GOD was immobilized on all of the tested surfaces and the highest glucose detection sensitivity of 60 μ M was obtained when the Pt nanowires were modified by PVA. Moreover, the sensors also showed very high current response when the Pt nanowires were modified with the cysteamine SAM.*

Keywords: *Platinum nanowires, deposition and etching under angle, surface modification, glucose oxidase, glucose detection.*

1. INTRODUCTION

Nanoscale devices based on nanowires have been realized for applications in electronics, optics, gas, and especially biomedical sensing [1–3]. One-dimensional structures such as nanowires are particularly compelling for electronic interconnects and biosensing applications due to their suitability for large-scale high-density integration and high sensitivity to surface interactions. Although nanowires have been fabricated by various methods [4–6], simple fabrication techniques which are not only easily addressed electrically, but also maintain reasonable costs

for practical application, are also highly desirable.

Surface properties are especially of concern because the interaction of any metal electrode with its environment mainly occurs at the surface, and also because of the dependence of the response on the surface state of the electrode. Many analytical applications, such as electron transfer reaction, preferential accumulation, or selective membrane permeation, can benefit from chemically modified electrodes [7–9]. Other important applications including electrochromic display devices, controlled release of drugs, electrosynthesis, corrosion protection, etc [10–14] can also benefit from the rational design of

electrode surfaces. Accordingly, deliberate modification of electrode surfaces can thus meet the needs of many electroanalytical problems [15, 16], and may form the basis for new analytical applications [17–19] and different sensing devices [20, 21]. One of the most important applications of platinum (Pt) nanowires electrode is glucose detection. To obtain a sensitive and realizable Pt-based glucose biosensor, one of the key steps is enzyme immobilization on the Pt surface for subsequent catalyst oxidation of glucose into sensible products. Up to now, various modification techniques have been applied in surface activation to immobilize the enzyme onto the Pt microwire electrode surface such as physical adsorption [22], entrapment [23], covalent binding [24], cross linking, etc.

In this paper we present a new fabrication technique that only uses conventional techniques of microtechnology such as microlithography, thin-film deposition and directional ion beam etching, named deposition and etching under angles (DEA). The DEA technique can make very narrow, wafer-scale length platinum (Pt) nanowires. Pt nanowire arrays, with wire width down to 30 nm and wire length up to several millimeters, have been realized on silicon chips. Additionally, the fabricated Pt nanowires are realized with electrical contact paths, and thus are ready for further electrical measurement and applications. Fabricated Pt nanowires electrodes were immobilized with GOD by using different techniques to investigate three

generations of glucose sensor. In the first generation, enzymes were immobilized via membrane silica–gel (SiO_2 + gelatin). This membrane creates a flexible matrix, negligible swelling in aqueous solution and thermal stability on the electrode [25]. In the second generation, GODs were immobilized through a polyvinyl alcohol (PVA) layer and a Prussian blue (PB) mediator. In the last generation, GOD immobilization influence was also studied for the self-assembled monolayers (SAMs) of cysteamine onto the platinum surface [26]. In addition, the performance of the glucose biosensors, including the response time, enzymatic sensitivity and device durability, are reported.

2. METHODS

2.1. Chemicals and apparatus

D-glucose and glucose oxidase (GOx, EC 1.1.3.4, 172 000 units g^{-1} from *Aspergillus niger*) were purchased from Sigma Aldrich. Gelatin (Merck) solution was dissolved in 0.05M acetate buffer pH 5.5 (CH_3COOH , CH_3COONa) and stirred for 1 h at 70°C . 25 wt% glutaraldehyde solution and tetraethyl ortho-silicate (TEOS) were purchased from Merck. SiO_2 solution was prepared by mixing 0.2 ml TEOS with 20 mL Ethanol 100%, 0.3 ml NH_4OH , 0.3 ml H_2O and 1 ml HCl in a glass vial. Then the homogeneous solution was obtained by stirring the solution at 80°C for 7 h. Polyvinylalcohol (PVA), cysteamine and aminopropyl triethoxysilane were obtained from

Sigma, while potassiumferricyanide ($\text{K}_3\text{Fe}(\text{CN})_6$) and ferricchloride (FeCl_3) were obtained from Aldrich. A 0.05 M phosphate buffer (PBS) solution was prepared using Na_2HPO_4 and KH_2PO_4 . All solutions were filtered through a syringe cellulose acetate ($0.22\ \mu\text{m}$) before use. Double distilled deionized water was used throughout the experiment.

All electrochemical measurements were carried out on Potentiostat/Galvanostat EG&G273A in a three-electrode conventional cell including the gold nanowires chip as working electrode, a platinum rod 0.5 mm diameter was used as a counter electrode, and a Ag/AgCl electrode as reference. All measurements were carried out under room temperature.

2.2. Fabrication of Pt nanowires by the DEA technique

The new fabrication process that has been developed and allows the fabrication of long and narrow Pt nanowires is shown schematically in figure 1. Briefly, a layer of 1000 nm silicon dioxide (SiO_2) is grown on a 4 inch, (100) silicon wafer by means of wet oxidation. Conventional microlithography is then carried out to define patterns on the wafer, followed by isotropic etching of SiO_2 for 1 min in a buffered oxide etching (BHF) solution. This isotropic etching creates an under-etching or nano-spacer with width about 65–70 nm below the photoresist layer.

Layers of 40 nm platinum/5 nm chromium are then deposited by an E-beam evaporator

with an inclined angle of 30° on the surface of the patterned wafer. The typical evaporation rate is $1\ \text{\AA}\ \text{s}^{-1}$ for both Cr and Pt. As the result of inclined deposition, a small part of the Pt/Cr is deposited into the nano-spacer or hidden below the photoresist film. In our work, Cr is used as an adhesive material for deposition of Pt film, and the width of the hidden metallic part depends on several parameters, such as the dimensions of the nano-spacer and the inclined evaporation angle.

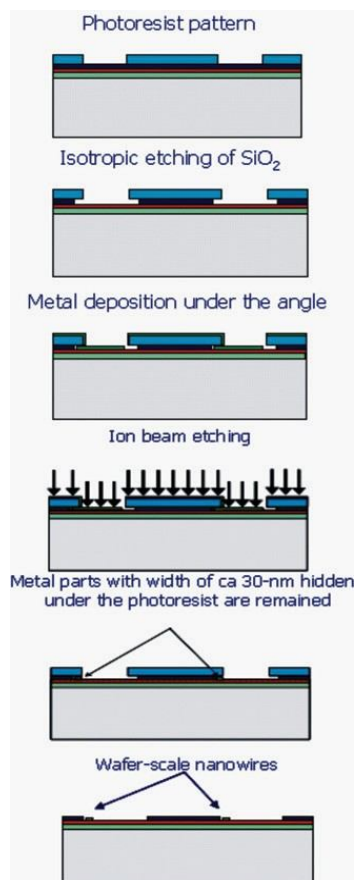


Figure 1. DEA fabrication process to make wafer-scale Pt nanowire using only conventional microfabrication techniques.

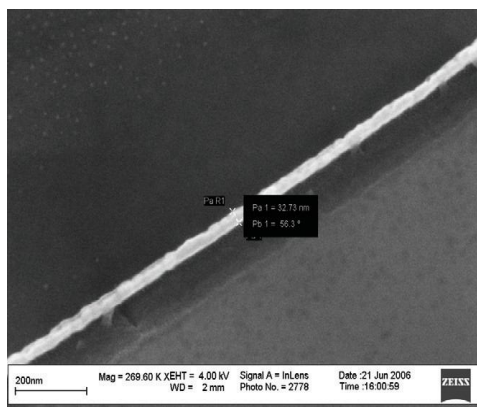


Figure 2. High resolution SEM image of the DEA fabricated Pt nanowire with width of about 32 ± 5 nm.

Subsequently, argon (Ar) ion beam etching (IBE) is carried out to remove the deposited Pt/Cr film from the silicon wafer. However, the metallic parts that are hidden below the photoresist film are not being reached by the Ar ion flux. Thus they are not etched, and remain along and below the photoresist pattern. The remaining metallic parts have a width of about 30 nm, therefore forming the metallic nanowires, which are Pt/Cr nanowires in the current work. The photoresist layer is subsequently removed in a hot acetone solution to reveal the Pt/Cr nanowires (figure 2).

Lithography is then carried out, followed by metallization to create macro contact pads for the individual Pt/Cr nanowires. Finally, the wafer containing Pt/Cr nanowires is diced into small chips with typical size of 7×7 mm (fig. 3). Each diced chip has 10 Pt nanowires several micrometers in length and about 40 nm in width, and any one of the realized Pt nanowires is individually electrically addressed through

its contact pads at both ends (see the inset of fig. 3).

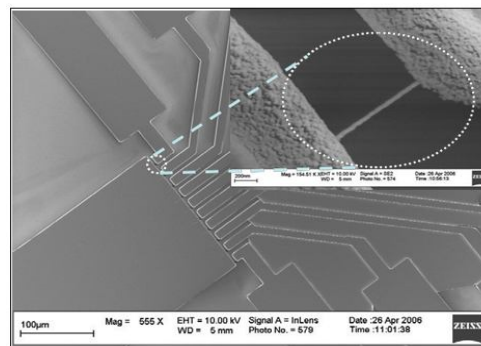


Figure 3. A diced chip contains an array of Pt nanowires. The inset image shows individually electrically addressed Pt nanowires, thus making the nanowires ready for measurement.

2.3. Preparation of enzyme electrode on different modified surface of Pt nanowire

Pt nanowires chips were immersed in dichloromethane, propanol, acetone and deionized water (DI) for 5 min, respectively. Then the samples were dried with blown nitrogen and cleaned by using oxygen plasma (power of 250 W for 6–7 min).

Then it was electrochemically scanned repeatedly until the voltammogram characteristic was obtained. In the first generation of glucose sensor, the cleaned electrode was immersed into the compound of 1 ml gelatin-SiO₂ (3:1 v/v mixture of concentrated gelatin, SiO₂ stirred in 2 h) and 0.5 ml GOD (5 mg/ml of acetate buffer, pH 5.5) solution. Afterwards, the electrode was dried at 4°C and washed with DI water before being used for glucose detection. In the next experiment, the electrode was reduced by scanning it in 0.001 M H₂ SO₄. Then it was

soaked into an ethanol solution containing cysteamine 0.25 M at 4°C for 12 h. Afterwards, this electrode was immersed into glutaraldehyde (GAD) solution (5 mg ml⁻¹ of PBS buffer) for 2 h. Finally, the modified electrode was soaked in GOD solution to bind the free enzyme from the solution onto the platinum surface.

Following the study of enzyme immobilization, PB film was electrodeposited onto the Pt nanowire surface by scanning the solution of 30 mM K₃Fe(CN)₆, 40 mM FeCl₃ and 1 M KCl:1 M HCl solution. The potential was scanned between -0.2 V to 0.8 V with 50 mV s⁻¹ in scan rate. In order to firm the PB mediator, we scanned it in 1 M KCl between -0.2 and 0.8 V. Then the modified electrode was immersed successively in PVA (5 mg ml⁻¹) solution and aminopropyltriethoxylane 90% for 30 mins and GOD for 3 h. In these experiments, the electrode was dried before dipping into each solution. All enzyme electrodes were kept at 4°C until use.

3. RESULT AND DISCUSSION

3.1. Fabrication of the Pt/Cr nanowires

Figure 2 shows a high resolution scanning electron microscopy (HR: SEM) image of the fabricated Pt nanowire. It can be seen that the realized nanowire has a width of about 32 ± 5 nm. Moreover, it is straight and with a smooth surface. The obtained results prove that we have successfully developed a new fabrication method that only utilizes conventional, thus inexpensive, microfabrication techniques to

realize very small Pt nanowires with good morphology.

Moreover, by adjusting several processing parameters such as the dimensions of the created nano-spacer (by varying the SiO₂ isotropic etching step) and inclining angles during metal film deposition and IBE etching, metallic nanowires with various widths can be obtained. However, in the current work we optimized process parameters to obtain Pt nanowires with width of around 35 nm, because wider nanowires may reduce the sensors' sensitivity while narrow ones may suffer the well-know problem of external noise.

Figure 3 shows a diced chip that contains an array of Pt nanowires, while the inset image shows that each nanowire from the array is individually electrically addressed. This allows the fabricated nanowires to easily be further connected to an outer electronics for detailed device measurement and applications.

3.2. Electrical characterization of the fabricated Pt nanowires

Figure 4 shows an I-V characterization of the 20 µm length Pt nanowires. It can be seen that the wires have good electrical characteristics with linear IV behavior of the bulk metal Pt. Moreover, the measurement results show a resistance of about 1540 ± 40 K for the fabricated Pt nanowire. This value is only about 30% higher than the value calculated using the bulk material.

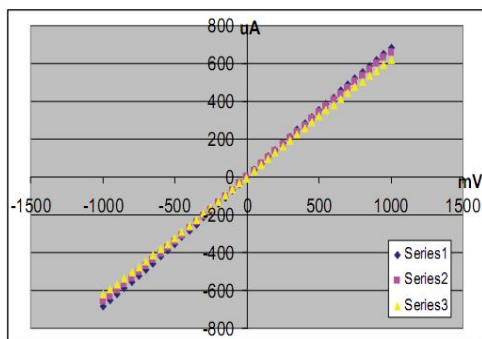


Figure 4. Current–voltage (I–V) curve, measured in ambient conditions, of the 20 μm length Pt.

3.3. Electrochemical characterization of Pt nanowire

Cyclic voltammograms (CVs) were performed in glucose solution in PBS buffer and a variety of glucose concentrations in water to investigate the influence of electrolyte solution on the platinum electrode prior to the immobilization process. We found that the current response of the electrode did not

3.4. Effect of pH on enzyme electrode

The influence of pH buffer solution on glucose detection has been studied by several authors [7–10]. Investigation of the effect of pH value on the performance of the glucose sensor is very important because the activity of immobilized GOD is pH dependent [8]. In our work, the pH dependence of a modified electrode by PVA compound and PB mediator was evaluated over the pH range from 5.6 to 8.4. When the pH of the buffer was very low or very high, the GOD electrode exhibited low current response to glucose. An optimum response current was observed at a pH value of 7.2.

appropriately change when increasing the concentration of the PBS at 0.2–0.8 V. In contrast, when the concentration of glucose in water increased, then all peak currents decreased immediately (figure 5). That phenomenon proves that all of these elements on the electrolyte did not react together but they react with the bare Pt nanowire surface.

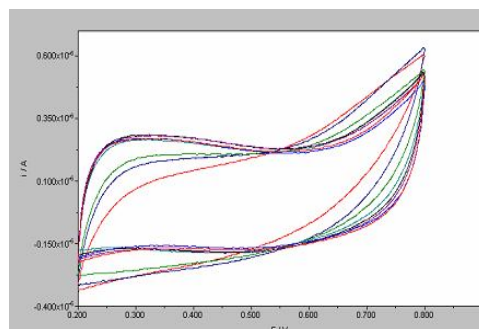


Figure 5. Current–voltage (C–V) characteristics of Pt nanowires electrode in glucose solution in various concentrations at 200 mVs⁻¹. From inside to outside 0, 2.5, 5, 10, 20 and 40 mM.

3.5. Cyclic voltammograms of enzyme electrodes

The response current of glucose on three types of biosensors was recorded and is shown in figure 6 with a potential scan rate of 100 mVs. The results show that all enzyme electrodes have high electron transfer efficiencies. We observed that with an increase in glucose concentration the redox current increased monotonously at a potential higher than 0.4 V and it just became stable only when the applied voltage was higher than 0.6 V. In contrast, the CV curve of a gel-SiO₂ modified electrode had an unstable current, and the applied voltage was higher than 0.7 V because of the influence of the oxygen concentration in

electrochemical solution. This is important information for applying different immobilization membranes and the mediator. Moreover, we also found that the oxidation current or reduction current increased linearly with the concentration of glucose, and this important result is reported in detail in the next section.

3.6. Amperometric response of glucose sensor

Figure 7 shows the dependence on glucose concentration (0–16 mM) of the CV curves of the electrodes modified by the three immobilizing methods. Obviously, the gelatin/SiO₂ modified Pt had the lowest response current and corresponding coefficient ($R^2 = 0.8335$). This indicated that this

modified surface had very little immobilized enzyme, thus little H₂O₂ was gained in the reaction with glucose. Samples with PB as the electron transfer mediator in PVA-PB-Pt obtained glucose detection sensitivities at 60 μ M ($R^2 = 0.955$). However, the highest response current was obtained with the electrode modified with the self-assembled layer of cysteamine ($R^2 = 0.9212$). The modifying chemicals in this case might create a suitable microenvironment that benefits the exposition of the enzyme activity center and increases the response current. This study suggests that the enzyme immobilized on different surfaces has distinct effectiveness, thus a stable and sensitive glucose sensor may need a combination of the above immobilizing methods.

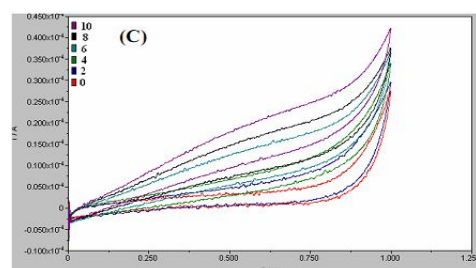
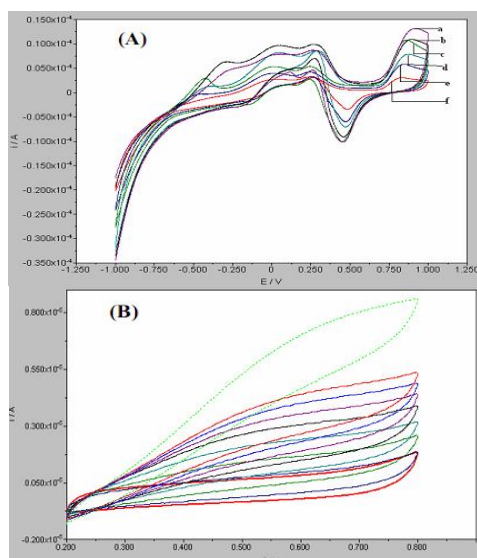


Figure 6. CV curves of different concentrations of glucose measured by (A) GOD-gelatin/SiO₂-Pt electrode, from down to up 0, 2, 4, 6, 8 and 16 mM; (B) GOD-PVA/PB-Pt electrode, from down to up 0, 2, 4, 8 and 12 mM; (C) GOD-cysteamine-Pt electrode, from down to up 0, 2, 4, 6, 8 and 10 mM.

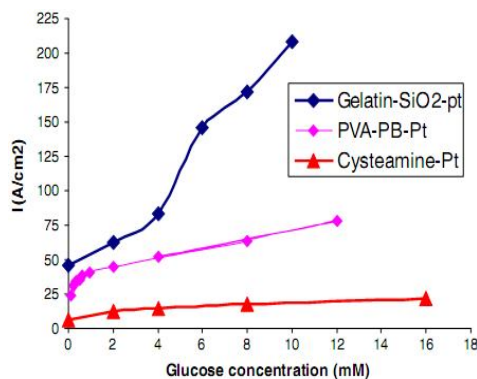


Figure 7. The response current of a glucose sensor modified by different immobilized surfaces of Pt nanowire at a potential of 0.6 V.

3.7. Reproducibility and stability of the glucose sensor

The PVA-GOD modified Pt nanowire electrodes were prepared under the same conditions described above for detecting 3 mM

4. CONCLUSION

A new fabrication process, DEA, has been developed that allows successful and inexpensive fabrication of narrow but long Pt nanowires. The fabricated Pt nanowire chips with appropriate dimensions and properties are then utilized to build a biosensor for accurate determination of the glucose concentration in aqueous solution.

The enzyme immobilization is influenced by linking chemical groups on different Pt surfaces, and the response current of the Pt nanowire based sensor is highly dependent on

glucose. The glucose sensor responses gradually decreased in the first 10 days, the activity remained constant at approximately 60% after 30 days, indicating good stability of the enzyme immobilized on the modified surfaces. Figure 8 shows the decrease in the current response, which is caused by leaking enzyme due to the loose links of the enzyme with the Pt surface after a considerable experiment period.

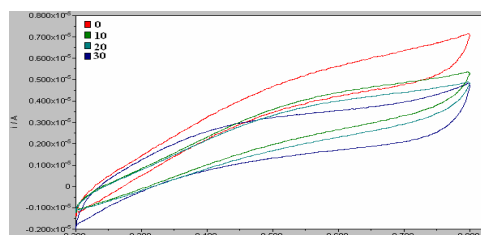


Figure 8. CV of enzyme electrode in 3 mM glucose solution at different times. From down to up 30, 20, 20 and 0 days, respectively.

the utilized surface modification methods. Our research results reveal that GOD immobilized on the Pt nanowires, which were previously modified by PVA with a PB mediator, gave the highest glucose detection sensitivities of about 60 μ M. The highest current response was achieved when the Pt nanowires were modified with the cysteamine SAM for subsequent binding of GOD. Furthermore, the stability and catalyst activity of the GOD were retained at about 60% after a store period of 30 days.

CHẾ TẠO VÀ HOẠT HÓA BỀ MẶT SỢI NANO PLATIN ỨNG DỤNG TRONG ĐỊNH LƯỢNG GLUCOSE

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TÓM TẮT: Trong bài báo này, một phương pháp mới - lắng đọng và ăn mòn dưới góc nghiêng (Deposition and Etching under Angle - DEA) được nghiên cứu để chế tạo số lượng lớn chip sợi nano platin ở qui mô cả phiến và các chip chế tạo ra có thể sử dụng ngay trong các đo đạc thực nghiệm tiếp theo. Phương pháp chế tạo này sử dụng những kỹ thuật cơ bản của công nghệ chế tạo micro thông thường, như là quang khắc quang học, lắng đọng màng mỏng và ăn mòn ion ở qui mô cả phiến, để chế tạo các dây sợi nano platin trên phiến silic với lớp cách điện silic điôxít. Chip sợi nano platin được chế tạo bên trên sau đó được hoạt hóa bằng các loại hóa chất khác nhau như là hỗn hợp của gel gelatin với SiO₂, polyvinyl ancol (PVA) và lớp đơn phân tử tự lắp ghép cysteamine (SAM). Sau đó, enzyme glucose oxidase được gắn lên các chip đã được hoạt hóa bề mặt để xác định nồng độ glucose trong dung dịch nước. Kết quả khảo sát chỉ ra rằng enzyme glucose oxidase (GOD) đã được gắn kết thành công lên bề mặt sợi platin được hoạt hóa bằng các phương pháp nêu trên và độ nhạy cao nhất của các chip với dung dịch glucose là 60 μ M với chip được hoạt hóa bằng phương pháp polyme hóa sử dụng polyvinyl ancol (PVA) với màng trung chuyển điện tử là Prussian Blue (PB). Bên cạnh đó, đối với chip được hoạt hóa bằng phương pháp lớp đơn phân tử tự lắp ghép cysteamine thì cường độ dòng đo được có giá trị lớn nhất.

Từ khóa: sợi nano Platin, phương pháp lắng đọng và ăn mòn dưới góc nghiêng (DEA), hoạt hóa bề mặt, glucose oxidase, phát hiện glucose.

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